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(FILE 'CAPREVIEWS' ENTERED AT 07:58:00 ON 10 JUL 95)
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FILE 'HCA' ENTERED AT 08:03:17 ON 10 JUL 95

L1 E KOHN D/AU
35 S E3-6
L2 E KOHN DONALD/AU
21 S E3-4
E BLAESE, M/AU
E BLAESE M/AU
L3 9 S E3-5
E MULLEN C/AU
L4 14 S E3-6 OR E11
E MOEN R/AU
L5 25 S E4-6 OR E9-11
L6 100 S L1 OR L2 OR L3 OR L4 OR L5
SAVE L6 TEMP MILNE/A
L7 854 S ((RETRO (L) VIR?) OR RETROVIR?) (L) VECTOR#
L8 2267 S ADENOSINE DEAMINASE#
L9 48 S IMMUNE DEFICIENCY (L) COMBIN?
L10 21 S SEVERE COMBINED IMMUNE DEFICIENCY
L11 245 S CD34
L12 18 S L6 AND (L7 OR L9 OR L10 OR L11)
L13 8 S L6 AND L8
L14 21 S L13 OR L12 *in vendor search*
L15 5454 S RETROVIR? OR RETRO (L) VIR?
L16 2 S L15 AND L8 AND L11
L17 75 S L15 AND L8
L18 15 S L15 AND L11

=> fil hca

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=> d bib ab 114 1-21;d bib abs hitind 116 1-2

L14 ANSWER 1 OF 21 HCA COPYRIGHT 1995 ACS
AN 123:7845 HCA
TI Retrovirally marked **CD34**-enriched peripheral blood and
bone marrow cells contribute to long-term engraftment after
autologous transplantation
AU Dunbar, Cynthia; Cottler-Fox, Michelle; O'Shaughnessy, Joyce A.;
Doren, Sandra; Carter, Charles; Brenson, Ronald; Brown, Sherri;
Moen, Robert C.; Greenblatt, Jay; et al.
CS Hematol. Branch, Natl. Institutes Health, Bethesda, MD, USA
SO Blood (1995), 85(11), 3048-57
CODEN: BLOOAW; ISSN: 0006-4971

DT Journal
 LA English
 AB A preliminary human autologous transplantation study of retroviral gene transfer to bone marrow (BM) and peripheral blood (PB)-derived CD34-enriched cells is described. Eleven patients with multiple myeloma or breast cancer had cyclophosphamide and filgrastim-mobilized PB cells CD34-enriched and transduced with a retroviral marking vector contg. the neomycin resistance gene, and CD34-enriched BM cells transduced with a second marking vector also contg. a neomycin resistance gene. After high-dose conditioning therapy, both transduced cell populations were reinfused and patients were followed over time for the presence of the marker gene and any adverse effects related to the gene-transfer procedure. All 10 evaluable patients had the marker gene detected at the time of engraftment, and 3 of 9 patients had persistence of the marker gene for greater than 18 mo post-transplantation. The marker gene was detected in multiple lineages, including granulocytes, T cells, and B cells. The source of the marking was both the transduced PB graft and the BM graft, with a suggestion of better long-term marking originating from the PB graft. The steady-state levels of marking were low, with only 1:1000 to 1:10,000 cells pos. There was no toxicity noted, and patients did not develop detectable replication-competent helper virus at any time post-transplantation. These results suggest that mobilized PB cells may be preferable to BM for gene therapy applications and that progeny of mobilized peripheral blood cells can contribute long-term to engraftment of multiple lineages.

L14 ANSWER 2 OF 21 HCA COPYRIGHT 1995 ACS
 AN 122:129910 HCA
 TI Multiple modifications in cis elements of the long terminal repeat of **retroviral vectors** lead to increased expression and decreased DNA methylation in embryonic carcinoma cells
 AU Challita, Pia-Maria; Skelton, Dianne; El-Khoueiry, Anthony; Yu, Xiao-Jin; Weinberg, Kenneth; **Kohn, Donald B.**
 CS Dep. Microbiol. Pediatrics, Univ. Southern California Sch. Med., Los Angeles, CA, USA
 SO J. Virol. (1995), 69(2), 748-55
 CODEN: JOVIAM; ISSN: 0022-538X
 DT Journal
 LA English
 AB Infection by murine retroviruses in embryonic carcinoma (EC) and embryonic stem cells is highly restricted. The transcriptional unit of the Moloney murine leukemic virus (MoMuLV) long terminal repeat (LTR) is inactive in EC and embryonic stem cells in assocn. with increased proviral methylation. In this study, expression in F9 EC cells was achieved from novel retroviral vectors contg. three modifications in the MoMuLV-based retroviral vector: presence of the myeloproliferative sarcoma virus LTR, substitution of the primer binding site, and either deletion of a neg. control region at the 5' end of the LTR or insertion of a demethylating sequence. The authors conclude that inhibition of expression from the MoMuLV LTR in EC cells is mediated through the additive effects of multiple cis-acting elements affecting the state of methylation of the

provirus.

L14 ANSWER 3 OF 21 HCA COPYRIGHT 1995 ACS

AN 122:98089 HCA

TI **Retroviral vectors** containing chimeric promoter/enhancer elements exhibit cell-type-specific gene expression

AU Couture, Larry A.; Mullen, Craig A.; Morgan, Richard A.

CS National Heart, Lung, and Blood Institute, Bethesda, MD, 20892, USA

SO Hum. Gene Ther. (1994), 5(6), 667-77

CODEN: HGTHE3; ISSN: 1043-0342

DT Journal

LA English

AB Retroviral vectors were constructed in which the U3 promoter/enhancer of Moloney murine leukemia (MoMLV) was replaced by the corresponding region from five related murine retroviruses - AKR murine leukemia virus (AKV), Harvey murine sarcoma virus (HaMSV), myeloproliferative sarcoma virus (MPSV), SL3-3, and the NZB-xenotropic virus (Xeno). In these vectors the chimeric long terminal repeat (chLTR) drives the expression of the chloramphenicol acetyl transferase (CAT) reporter gene that is followed by an internal SV40 virus early region promoter linked to the neomycin phosphotransferase II (NEO) gene. As an initial measure of the relative promoter/enhancer strength of the chLTR vectors, the murine NIH-3T3 cell line and the human JURKAT cell lines were transfected and assayed for CAT reporter activity. Relative to the MoMLV vector, the HaMSV construct was the most active in NIH-3T3 cells whereas the SL3-3 vector displayed the greatest activity in JURKAT cells. Retroviral vector producer cell populations and cell clones were established for each chLTR vector, and all were capable of yielding high vector titers (>10⁵ G418R cfu/mL on NIH-3T3). Supernatant from these cells was used to transduce both mouse and human cell lines and primary cells. In NIH-3T3 cells and two murine fibrosarcoma cell lines, the HaMSV chLTR vector was slightly more active than the MoMLV chLTR vector. In the human HepG2 and HeLa cell lines, the MPSV chLTR vector was the most active. Data from the human JURKAT T-cell line and a T-cell line derived from an ADA-deficient severe combined immunodeficiency (SCID) patient demonstrate that the SL3-3 chLTR is the most active in these lymphoid cell lines. The greatest difference in the comparison of the different chLTR vectors was obsd. in primary human umbilical vein endothelial cells, where the MoMLV vector produced up to 100 times more CAT activity than the SL3-3 vector. These data suggest that the use of specific promoter/enhancer elements may lead to higher levels of gene expression following retroviral-mediated gene transfer into specific cell types and these observations may be useful in the design of human gene therapy expts.

L14 ANSWER 4 OF 21 HCA COPYRIGHT 1995 ACS

AN 122:96039 HCA

TI Development of live tumor vaccines using **retroviral vectors** for transfer of suicide genes and cytokines

AU Mullen, Craig A.

CS Diagnosis and Centers, National Cancer Institute, Bethesda, MD, USA

SO Contrib. Oncol. (1994), 46(CYTOKINES IN CANCER THERAPY), 260-8

CODEN: COONEV; ISSN: 0250-3220

DT Journal

LA English

AB Recent work with suicide genes and cytokines indicates the following:.. Animals can be treated with large doses of the prodrug 5-fluorocytosine (5-FC) without serious toxicity. Live cytosine deaminase (CD)+ tumors can be eliminated in vivo by 5-FC treatment, suggesting that live attenuated strains of tumor can be produced. Animals rejecting CD+ tumors after 5-FC treatment develop immunity to wild-type tumor. Tumors modified to secrete IL-6 are rejected or grow more slowly than wild-type tumor. Animals rejecting IL-6-secreting tumor also develop immunity to wild-type tumor. Double transduction of tumor with CD and IL-6 genes allows delivery of much larger doses of cytokine-secreting tumor to the host. These results suggest that live attenuated tumor vaccines can be produced with gene transfer techniques and that work to find optimal combinations of genes and immunization schedules is justified.

L14 ANSWER 5 OF 21 HCA COPYRIGHT 1995 ACS

AN 122:45771 HCA

TI Inhibition of HIV-1 in human T-lymphocytes by retrovirally transduced anti-tat and rev hammerhead ribozymes

AU Zhou, Chen; Bahner, Ingrid C.; Larson, Garry P.; Zaia, John A.; Rossi, John J.; **Kohn, Donald B.**

CS Division of Research Immunology and Bone Marrow Transplantation, Childrens Hospital Los Angeles, Departments of Pediatrics and Microbiology, University of Southern California School of Medicine, Los Angeles, CA, 90027, USA

SO Gene (1994), 149(1), 33-9

CODEN: GENED6; ISSN: 0378-1119

DT Journal

LA English

AB Gene therapy for AIDS requires the identification of genes which effectively inhibit HIV-1 replication coupled to an efficient vector system for gene delivery and expression. Hammerhead ribozymes are RNA mols. capable of catalytic cleavage of complementary RNA mols. Ribozymes targeted against two portions of the HIV-1 genome were designed to cleave HIV RNA in the tat gene (TAT) or in a common exon for tat and rev (TR). The ribozymes were cloned into the LN (LTR-neomycin) retroviral vector plasmids and expressed as part of viral LTR-driven transcripts. The vectors were packaged as amphotropic virions and used to transduce human T-lymphocytes. Expression of the vector transcripts contg. the ribozyme sequences was readily detected by Northern blot anal. of the transduced T cells. The T-lymphocytes expressing the anti-HIV-1 ribozymes showed resistance to HIV-1 replication. In contrast, cells expressing mutant ribozymes, contg. substitutions of a key nucleotide in the catalytic domain which cripples the cleavage activity of the ribozymes, supported replication of HIV-1, demonstrating that the functional ribozymes were cleaving the target RNAs. These studies demonstrate that retrovirally transduced ribozymes included in long, multifunctional transcripts, can inhibit HIV replication in human T-lymphocytes. The ribozyme and expression strategies described here should be useful for the gene therapy of AIDS by conferring resistance to HIV-1 replication on cells derived from transduced

hematopoietic stem cells.

L14 ANSWER 6 OF 21 HCA COPYRIGHT 1995 ACS

AN 120:316888 HCA

TI Lack of expression from a **retroviral vector** after transduction of murine hematopoietic stem cells is associated with methylation in vivo

AU Challita, Pia Maria; **Kohn, Donald B.**

CS Sch. Med., Univ. Southern California, Los Angeles, CA, 90027, USA

SO Proc. Natl. Acad. Sci. U. S. A. (1994), 91(7), 2567-71

CODEN: PNASA6; ISSN: 0027-8424

DT Journal

LA English

AB The authors describe studies of gene transfer and expression of the human glucocerebrosidase cDNA by a Moloney murine leukemia virus (MoMuLV)-based retroviral vector in a murine gene transfer/bone marrow transplant (BMT) model. Pluripotent hematopoietic stem cells (HSCs) were assayed as the colony-forming units, spleen (CFU-S) generated after serial transplantation. Transcriptional expression from the MoMuLV long-terminal repeat (LTR) was detected at a high level in the primary (1.degree.) CFU-S and tissues of reconstituted BMT recipients. However, the authors obsd. transcriptional inactivity of the proviral MoMuLV-LTR in >90% of the secondary (2.degree.) CFU-S and in 100% of the tertiary (3.degree.) CFU-S examd. The authors have compared the methylation status of the provirus in the 1.degree. CFU-S, which show strong vector expression, to that of the transcriptionally inactive provirus in the 2.degree. and 3.degree. CFU-S by Southern blot anal. using the methylation-sensitive restriction enzyme Sma I. The studies demonstrated a 3- to 4-fold increase in methylation of the SmaI site in the proviral LTR of 2.degree. and 3.degree. CFU-S compared to the transcriptionally active 1.degree. CFU-S. These observations may have important implications for future clin. applications of retroviral-mediated gene transfer into HSCs, where persistent gene expression would be needed for an enduring therapeutic effect.

L14 ANSWER 7 OF 21 HCA COPYRIGHT 1995 ACS

AN 120:126753 HCA

TI Growth factors increase amphotropic retrovirus binding to human **CD34+** bone marrow progenitor cells

AU Crooks, Gay M.; **Kohn, Donald B.**

CS Div. Res. Immunol., Child. Hosp. Los Angeles, Los Angeles, CA, USA

SO Blood (1993), 82(11), 3290-7

CODEN: BLOOAW; ISSN: 0006-4971

DT Journal

LA English

AB Gene transfer into human cells using murine amphotropic retroviral vectors is the basic technique used in most current gene therapy studies. The identity of the cell surface receptor for the amphotropic envelope remains unknown and thus its importance in gene transfer is poorly understood. The authors have measured specific retrovirus binding to cells to study amphotropic virus receptor regulation in human CD34+ CD38- human hematopoietic cells. The rat monoclonal antibody 83A25 recognizes an epitope common to the envelope glycoprotein of all classes of Mononey murine leukemia

virus. Indirect fluorescent labeling of 83A25 allows flow cytometric anal. of specific virus-cell interactions and is an indirect measure of specific receptors. Using this assay, amphotropic virus binding to fresh CD34+ cells were minimal. However, when CD34+ cells were cultured with or without growth factors for 4 days, specific binding of amphotropic retrovirus was readily shown. Inclusion of interleukin-3 (IL-3), IL-6, and Steel factor in cultures increased the fluorescence assocd. with amphotropic virus binding by 2-4 fold (mean fold increase 2.7 ± 0.84). Virus binding to CD34+ CD38- cells was shown only in those cells cultured in IL-3, IL-6, and Steel factor. These results suggest that certain cytokines may cause an increase in the no. and/or affinity of amphotropic receptors on primitive human hematopoietic cells. Upregulation of viral receptor expression may be one of the mechanisms by which cytokines enhance gene transfer into primitive BM cells.

L14 ANSWER 8 OF 21 HCA COPYRIGHT 1995 ACS

AN 120:23215 HCA

TI T lymphocyte ontogeny in **adenosine deaminase**
-deficient **severe combined immune**

deficiency after treatment with polyethylene glycol-modified
adenosine deaminase

AU Weinberg, Kenneth; Hershfield, Michael S.; Bastian, John; **Kohn, Donald**; Sender, Leonard; Parkman, Robertson; Lenarsky, Carl

CS Sch. Med., Univ. South. California, Los Angeles, CA, 90027, USA

SO J. Clin. Invest. (1993), 92(2), 596-602

CODEN: JCINAO; ISSN: 0021-9738

DT Journal

LA English

AB Adenosine deaminase (ADA) deficiency causes severe combined immune deficiency (SCID) by interfering with the metab. of deoxyadenosine, which is toxic to T lymphocytes at all stages of differentiation. Enzyme replacement with polyethylene glycol-modified ADA (PEG-ADA) has been previously shown to correct deoxyadenosine metab. and improve mitogen-induced T lymphocyte proliferation. The authors studied the biochem. and immunol. effects of PEG-ADA in two infants with ADA-deficient SCID. While in a catabolic state, higher doses of PEG-ADA than previously described were required to normalize deoxyadenosine metab. After biochem. improvement, the patients recovered immune function in a pattern similar to that obsd. in normal thymic ontogeny and in patients with immunol. reconstitution after bone marrow transplantation. Immune reconstitution was marked by the sequential appearance in the peripheral blood of phenotypic T lymphocytes corresponding to successive stages of thymic differentiation. Functional reconstitution was marked by the successive appearance of mitogen responses dependent on exogenous in vitro IL-2, mitogen responses not requiring exogenous IL-2, antigen-specific responses dependent on exogenous IL-2, and finally, antigen-specific responses not requiring exogenous IL-2. Natural killer function was tested in one patient and normalized with PEG-ADA therapy. Optimal PEG-ADA therapy appears to normalize thymic differentiation in ADA-deficient SCID, resulting in normal antigen-specific immune function.

L14 ANSWER 9 OF 21 HCA COPYRIGHT 1995 ACS

AN 119:87556 HCA

TI Comparison of trans-dominant inhibitory mutant human immunodeficiency virus type 1 genes expressed by **retroviral vectors** in human T lymphocytes

AU Bahner, Ingrid; Zhou, Chen; Yu, Xiao-Jin; Hao, Qian-Lin; Guatelli, John C.; **Kohn, Donald B.**

CS Div. Res. Immunol./Bone Marrow Transplant., Child. Hosp., Los Angeles, CA, 90027, USA

SO J. Virol. (1993), 67(6), 3199-207.

CODEN: JOVIAM; ISSN: 0022-538X

DT Journal

LA English

AB Trans-dominant inhibitory mutant versions of the human immunodeficiency virus type 1 (HIV-1) regulatory genes tat and rev have previously been described. The authors have constructed a series of retroviral vectors to transduce these genes and compare their inhibitory activities. The inhibitory activities were measured with transient transfection assays by using a reporter which expresses an HIV-1 gag-Escherichia coli lacZ fusion protein with strict dependence on coexpression of both tat and rev. Addnl., the vectors were packaged as amphotropic virions and used to stably transduce human CEM T lymphocytes. The transduced CEM cells were challenged with HIV-1, and the effects of the mutant HIV-1 genes were detd. by measuring the levels of HIV-1 p24gag produced. A tat gene substituted at amino acid 41 (tatk41a) retained partial trans-activating activity and lacked inhibitory activity. A tat gene with a premature stop codon at amino acid 54 (tat54ter) showed moderate trans-dominant inhibition of the reporter plasmid but failed to significantly inhibit HIV-1 replication. The M10 rev mutant, with a 2-amino-acid substitution, showed strong trans-dominant inhibitory activity both in the reporter plasmid and in the HIV-1 infection assay. The greatest inhibition of HIV-1 growth was seen when M10 was expressed under the transcriptional control of a human cytomegalovirus promoter; slightly less inhibition was achieved when expression of M10 was controlled by the Moloney murine leukemia virus long terminal repeat, and minimal inhibition was seen when the HIV-1 long terminal repeat controlled the M10 gene. These results demonstrate the potential utility of retroviral vectors expressing trans-dominant inhibitory mutant HIV-1 genes for gene therapy approaches to AIDS.

L14 ANSWER 10 OF 21 HCA COPYRIGHT 1995 ACS

AN 118:52210 HCA

TI Retroviral-mediated transfer of the human glucocerebrosidase gene into cultured Gaucher bone marrow. [Erratum to document cited in CA117(15):143214t]

AU Nolte, Jan A.; Yu, Xiao Jin; Bahner, Ingrid; **Kohn, Donald B.**

CS Dep. Pediatr., Child. Hosp. Los Angeles, Los Angeles, CA, 90027, USA

SO J. Clin. Invest. (1992), 90(4), 1635

CODEN: JCINAO; ISSN: 0021-9738

DT Journal

LA English

AB An error in Figure 2 has been cor. The error was not reflected in the abstr. or the index entries.

L14 ANSWER 11 OF 21 HCA COPYRIGHT 1995 ACS

AN 117:143214 HCA

TI Retroviral-mediated transfer of the human glucocerebrosidase gene into cultured Gaucher bone marrow

AU Nolta, Jan A.; Yu, Xiao Jin; Bahner, Ingrid; **Kohn, Donald B.**

CS Dep. Pediatr., Child. Hosp. Los Angeles, Los Angeles, CA, 90027, USA

SO J. Clin. Invest. (1992), 90(2), 342-8

CODEN: JCINAO; ISSN: 0021-9738

DT Journal

LA English

AB Gaucher disease, a lysosomal glycolipid storage disorder, results from the genetic deficiency of an acidic glucosidase, glucocerebrosidase (GC). The beneficial effects of allogeneic bone marrow transplantation (BMT) for Gaucher disease suggest that GC gene transduction and the transplantation of autologous hematopoietic stem cells (gene therapy) may similarly alleviate symptoms. The authors constructed a retroviral vector, L-GC, produced by a clone of the amphotropic packaging cell line PA317, which transduces the normal human GC cDNA with high efficiency. Whole-marrow mononuclear cells and CD34-enriched cells from a 4-yr-old female with type 3 Gaucher disease were transduced by the L-GC vector and studied in long-term bone marrow culture (LTBMC). Prestimulation of marrow with IL-3 and IL-6, followed by co-cultivation with vector-producing fibroblasts, produced gene transfer into 40-45% of the hematopoietic progenitor cells. The levels of GC expression in progeny cells (primarily mature myelomonocytic) produced by the LTBMC were quant. analyzed by Northern blot, Western blot, and glucocerebrosidase enzyme assay. Normal levels of GC RNA, immunoreactive protein, and enzymic activity were detected throughout the duration of culture. These studies demonstrate that retroviral vectors can efficiently transfer the GC gene into long-lived hematopoietic progenitor cells from the bone marrow of patients with Gaucher disease and express physiol. relevant levels of GC enzyme activity.

L14 ANSWER 12 OF 21 HCA COPYRIGHT 1995 ACS

AN 112:133770 HCA

TI Expression of human glucocerebrosidase in murine long-term bone marrow cultures after **retroviral vector**-mediated transfer

AU Nolta, Jan A.; Sender, Leonard S.; Barranger, John A.; **Kohn, Donald B.**

CS Div. Res. Immunol., Child. Hosp. Los Angeles, Los Angeles, CA, USA

SO Blood (1990), 75(3), 787-97

CODEN: BLOOAW; ISSN: 0006-4971

DT Journal

LA English

AB A retroviral vector (N2-SV-GC) was constructed by inserting a normal human glucocerebrosidase (GC) cDNA under control of the SV40 early region promoter into the Moloney murine leukemia virus-derived N2 vector. N2-SV-GC produced human GC in murine 3T3 fibroblasts at levels in the range of the endogenous murine GC as detd. by enzymic assay and Western blot anal. The N2-SV-GC retroviral vector was used for studies of gene transduction of murine hematopoietic

progenitor cells (HPC). Infection of bone marrow cultured for 2 to 10 days in medium contg. hematopoietic growth factors was significantly more efficient than infection of freshly isolated marrow cells (24% to 32% G418-resistant CFU-GM 15%, resp.). The marrow infected by N2-SV-GC was maintained in long-term bone marrow culture (LTBMCE and had a stable level of G418-resistant HPC over 2 mo of serial assays. The human GC gene of the vector was persistently expressed in the nonadherent cell fraction of the murine LTBMCE as detd. by Northern blotting, Western blotting, and immunohistochem. staining using a monoclonal antibody specific for human GC. N2-SV-GC also expressed the human GC gene in day 12 CFU-S. LTBMCE represents a novel system for retroviral vector-mediated gene transduction of HPC and may accurately predict the activities of vectors in vivo.

- L14 ANSWER 13 OF 21 HCA COPYRIGHT 1995 ACS *photocopy* (AKM)
 AN 111:210043 HCA
 TI Transfer and expression of the human **adenosine**
****deaminase**** (ADA) gene in ADA-deficient human T lymphocytes
 with **retroviral vectors**
 AU **Kohn, Donald B.**; Kantoff, Philip; Zwiebel, James; Gilboa,
 Eli; Anderson, W. French; Blaese, R. Michael
 CS Metab. Branch, NCI, Bethesda, MD, 20892, USA
 SO UCLA Symp. Mol. Cell. Biol., New Ser. (1989), 87(Gene Transfer Gene
 Ther.), 365-74
 CODEN: USMBD6; ISSN: 0735-9543
 DT Journal
 LA English
 AB Transfer of the human ADA gene into an HTLV-1 transformed,
 ADA-deficient human T lymphocyte line (TJF-2) by a retroviral vector
 (SAX) was previously shown to produce normal levels of ADA activity.
 This report describes SAX infection of non-transformed T lymphocytes
 from ADA-deficient patients. This leads to increased levels of ADA
 activity in these primary T cells, similar to those produced in the
 transformed T line. To quantitate the rate of ADA gene transfer and
 expression by SAX, TJF-2 cells which were infected by SAX were
 cloned by limiting diln. Six out of 27 (22%) of the clones had
 acquired and were expressing the SAX vector. One to 3 copies of
 SAX/cell produced ADA activity in the range found in normal
 thymocytes and T lymphocytes. Similar vectors with other promoters
 were also highly active. Thus, these vectors are capable of
 efficient transfer and expression of the human ADA gene in
 ADA-deficient, human T cells.
- L14 ANSWER 14 OF 21 HCA COPYRIGHT 1995 ACS
 AN 111:188732 HCA
 TI Expression of the human glucocerebrosidase gene by
retrovirus vectors
 AU **Kohn, Donald B.**; Nolta, Jan A.; Hong, Chang Mu; Barranger,
 John A.
 CS Div. Res. Immunol. Bone Marrow Transplantat., Child. Hosp. Los
 Angeles, Los Angeles, CA, 90027, USA
 SO UCLA Symp. Mol. Cell. Biol., New Ser. (1989), 87(Gene Transfer Gene
 Ther.), 397-408
 CODEN: USMBD6; ISSN: 0735-9543

DT Journal
 LA English
 AB Gaucher disease, caused by glucocerebrosidase deficiency, may be a candidate for early trials of human gene therapy. A series of retrovirus vectors, which contain either a normal human glucocerebrosidase cDNA or a minigene with a 5' genomic glucocerebrosidase gene fragment fused to the 3' portion of the cDNA was constructed. These genes were under transcriptional control of either heterologous flanking region. Transfection or infection of the vectors into murine fibroblasts results in expression of glucocerebrosidase activity to levels equal to that of normal murine or human fibroblasts. The rank order of promoter activity is: glucocerebrosidase > SV40 > thymidine kinase. The conferred glucocerebrosidase activity is immunoprecipitable by a monoclonal antibody specific for the human enzyme. Western blots show the expressed protein is of the expected size range (59-66 kd). Southern blotting reveals that cells which express activity in the normal range contain a single intact copy of the vector. Retrovirus vectors are capable of high efficiency transduction of the human glucocerebrosidase gene and may be useful for clin. gene therapy.

L14 ANSWER 15 OF 21 HCA COPYRIGHT 1995 ACS

AN 111:110248 HCA

TI Retroviral-mediated gene transfer into hemopoietic cells

AU Eglitis, Martin A.; Kantoff, Philip W.; **Kohn, Donald B.**; Karson, Evelyn; **Moen, Robert C.**; Lothrop, Clinton D., Jr.; Blaese, R. Michael; Anderson, W. French

CS Lab. Mol. Hematol., NIH, Bethesda, MD, USA

SO Adv. Exp. Med. Biol. (1988), 241(Mol. Biol. Hemopoiesis), 19-27
 CODEN: AEMBAP; ISSN: 0065-2598

DT Journal

LA English

AB Retroviral vectors have provided a means for the introduction of functioning exogenous genes into the hematopoietic system of whole animals. Although these vectors are quite efficient in the mouse model, when applied to non-murine in vivo systems, the efficiency of gene transfer has diminished to impractical levels. Since in vivo analyses are expensive and time consuming, in vitro models have been developed to speed the evaluation of alternative protocols. Using in vitro colony assays, 3 approaches were evaluated for their ability to improve the infectivity of hematopoietic progenitor cells with retoviral vectors. Exogenously applied hematopoietic growth factors increased the proportion of hematopoietic colonies in vitro up to an av. of 5-fold. when alternative sources of progenitors, such as fetal cord blood, were used, improvements in infection efficiency were also obtained. Finally, evidence was acquired suggesting that xenotropic packaging of vectors also improved infection efficiency.

L14 ANSWER 16 OF 21 HCA COPYRIGHT 1995 ACS

AN 111:22025 HCA

TI Establishment and characterization of **adenosine deaminase**-deficient human T cell lines

AU **Kohn, Donald B.**; Mitsuya, Hiroaki; Ballow, Mark; Selegue, Jane E.; Barankiewicz, Jerzy; Cohen, Amos; Gelfand, Erwin; Anderson,

10% has been found 153 days after transplantation. Human bone marrow has also been treated with the N2 vector, resulting in 1-2% G418-resistant progenitors.

L14 ANSWER 18 OF 21 HCA COPYRIGHT 1995 ACS
 AN 107:212900 HCA
 TI Retroviral-mediated gene transfer into mammalian cells
 AU **Kohn, Donald B.**; Kantoff, Philip W.; Eglitis, Martin A.; McLachlin, Jeanne R.; **Moen, Robert C.**; Karson, Evelyn; Zwiebel, James A.; Nienhuis, Arthur; Karlsson, Stefan; et al.
 CS Metab. Branch, Natl. Cancer Inst., Bethesda, MD, 20892, USA
 SO Blood Cells (1987), 13(1-2), 285-98
 CODEN: BLCEDD; ISSN: 0340-4684
 DT Journal
 LA English
 AB Retroviruses may be used as genetic vectors to transfer genes into mammalian cells with high efficiency. The N2 vector will transfer a functional bacterial gene for neomycin resistance (NeoR) into more than 80% of mouse spleen foci. A deriv. of the N2 vector was constructed to study transfer and expression of the human gene for adenosine deaminase (ADA) in mammalian lymphoid and hematopoietic stem cells. This vector, termed SAX, contains the human ADA cDNA with an SV40 promoter in addn. to the NeoR gene. The SAX vector was found to efficiently transfer and express the ADA gene in an ADA-deficient human T-cell line. Gene transfer by SAX using an autologous nonhuman primate bone marrow transplant model resulted in expression of the human ADA gene in peripheral blood cells of treated animals. Human bone marrow treated with SAX produced 1-2% of colonies in vitro that were expressing the vector genes. Transfer of genes into circulating hematopoietic stem cells of fetal sheep in utero was most efficient; vector gene expression was evident in 20-40% of hematopoietic colonies. Therefore, retroviral vectors are capable of transferring functional genes into a wide variety of mammalian lymphoid and hematopoietic cells. Such vectors may be useful for clin. trials of gene therapy, i.e., the correction of genetic diseases by insertion of a normal gene into a patient's defective cells.

L14 ANSWER 19 OF 21 HCA COPYRIGHT 1995 ACS
 AN 107:91223 HCA
 TI Expression of human **adenosine deaminase** in nonhuman primates after retrovirus-mediated gene transfer
 AU Kantoff, Philip W.; Gillio, Alfred P.; McLachlin, Jeanne R.; Bordignon, Claudio; Eglitis, Martin A.; Kernan, Nancy A.; **Moen, Robert C.**; **Kohn, Donald B.**; Yu, Sheau Fung; et al.
 CS Lab. Mol. Hematol., Natl. Heart, Lung, Blood Inst., Bethesda, MD, 20892, USA
 SO J. Exp. Med. (1987), 166(1), 219-34
 CODEN: JEMEAV; ISSN: 0022-1007
 DT Journal
 LA English
 AB Primate bone marrow cells were infected with a retroviral vector carrying the genes for human adenosine deaminase (h-ADA) and bacterial neomycin resistance (neor). The infected cells were infused back into the lethally irradiated donor animals. Several

monkeys fully reconstituted and expressed the h-ADA and neor genes at low levels in their recirculating hematopoietic cells for short periods of time.

- L14 ANSWER 20 OF 21 HCA COPYRIGHT 1995 ACS
 AN 107:34292 HCA
 TI Retroviral-mediated gene transfer into hematopoietic cells
 AU Kantoff, Philip W.; Gillio, Al; McLachlin, Jeanne R.; Flake, Alan W.; Eglitis, Martin A.; Moen, Robert; Karlsson, Stefan; Kohn, Don B.; Karson, Evelyn; et al.
 CS Lab. Mol. Hematol., Natl. Heart, Lung, Blood Inst., Bethesda, MD, 20892, USA
 SO Trans. Assoc. Am. Physicians (1986), 99, 92-102
 CODEN: TAAPAI; ISSN: 0066-9458
 DT Journal
 LA English
 AB Protocols for gene transfer using bone marrow transplantation were developed in 2 large animal models, nonhumanprimates (rhesus monkey and cynomolgus monkey) and fetus of sheep. Retroviral vectors, N2 or SAX, carrying the transposon Tn5 neomycin resistance gene or the neomycin resistance gene and the human adenosine deaminase cDNA, resp., were transferred to isolated bone marrow cells by either co-cultivation with virus-producing fibroblasts or suspension in virus-contg. supernatant from virus-producing cells. After transfection, the bone marrow cells were washed and infused back into the donor animal. Reconstitution represented the full recovery of all images. Monkeys whose bone marrow cells were subjected to co-cultivation failed to fully reconstitute. However, after the supernatant infection protocol, the bone marrow cells showed full reconstitution and low but clearly detectable and reproducible levels of neomycin resistance and human adenosine deaminase gene activity. Circulating hematopoietic cells from a 100-day-old fetal lamb were removed in utero by exchange transfusion, infected with the N2 vector by the supernatant infection protocol, and infused back into the lamb. After the birth of the lamb, G418 resistant bone marrow cells were obtained from the infected animal but not from age-matched control lambs. When human bone marrow cells were transfected with the N2 vector by either the co-cultivation or the supernatant protocol, the levels of functional gene transfer was 1-2%. The potential utility of these methods in establishing a human gene therapy protocol is discussed.
- L14 ANSWER 21 OF 21 HCA COPYRIGHT 195 ACS
 AN 105:166144 HCA
 TI Correction of **adenosine deaminase** deficiency in cultured human T and B cells by retrovirus-mediated gene transfer
 AU Kantoff, Philip W.; Kohn, Donald B.; Mitsuya, Hiroaki; Armentano, Donna; Sieberg, Miri; Zwiebel, James A.; Eglitis, Martin A.; McLachlin, Jeanne R.; Wiginton, Dan A.; et al.
 CS Lab. Mol. Hematol., Natl. Heart, Lung, Blood Inst., Bethesda, MD, 20892, USA
 SO Proc. Natl. Acad. Sci. U. S. A. (1986), 83(17), 6563-7
 CODEN: PNASA6; ISSN: 0027-8424
 DT Journal
 LA English

AB A retroviral vector called SAX, contg. the cloned human cDNA for adenosine deaminase (ADA) [9026-93-1], was constructed and used to introduce the ADA gene into cultured T- and B-lymphocyte lines derived from patients with ADA deficiency. DNA anal. showed that the SAX vector was inserted intact into the T and B cells at .apprx.1 copy per cell. The treated cells produced the characteristic isoenzymes of human ADA at a level similar to normal T and B lymphocytes. It is known that ADA-deficient lymphocytes are unusually sensitive to high levels of 2'-deoxyadenosine, and this is the mechanism though to underlie the selective lymphocytotoxicity assocd. with ADA deficiency in vivo. Expression of the introduced ADA gene was sufficient to reverse the hypersensitivity of these genetically deficient lymphocytes to 2'-deoxyadenosine toxicity. These results support the suggestion that retroviral vector gene-delivery systems show promise for application to human gene therapy.

L16 ANSWER 1 OF 2 HCA COPYRIGHT 1995 ACS

AN 120:321192 HCA

TI Gene transfer into nonhuman primate CD34+CD11b-bone marrow progenitor cells capable of repopulating lymphoid and myeloid lineages

AU Van Beusechem, Victor W.; Bart-Baumeister, Julia A. K.; Bakx, Trudy A.; Kaptein, Leonie C. M.; Levinsky, Roland J.; Valerio, Dinko

CS TNO Med. Biol. Lab., Dep. Gene Ther., Rijswijk, Neth.

SO Hum. Gene Ther. (1994), 5(3), 295-305

CODEN: HGTHE3; ISSN: 1043-0342

DT Journal

LA English

AB The authors investigated whether rhesus monkey CD34+CD11b-hematopoietic stem cells can be transduced with recombinant retroviruses carrying the human adenosine deaminase (hADA) gene by co-cultivation with a virus-producing cell line. Following autologous transplantation, polymerase chain reaction (PCR) anal. on peripheral blood mononuclear cells and granulocytes showed that the hADA-retrovirus was present in approx. 0.1% of the cells for at least 400 days post transplantation in 2 monkeys. Bone marrow that was harvested 16 mo after transplantation carried ADA-overexpressing myeloid progenitor cells capable of in vitro colony formation. In addn., hADA activity could be demonstrated in T lymphocytes that were harvested 9 mo post transplantation. Thus, in vitro transduction of CD34+CD11b- cells led to long-term repopulation of the hematopoietic system with transduced cells of lymphoid and myeloid lineages expressing the hADA gene. To investigate whether infusion of virus-producing cells into a rhesus monkey undergoing autologous bone marrow transplantation could lead to in vivo transfer of the recombinant retrovirus, 1 monkey was infused with CD34+CD11b- bone marrow cells (BMC) and a large quantity of virus-producing cells. Few provirus-carrying cells could temporarily be detected in this animal. This shows that in vivo gene transfer into a regenerating hemopoietic system can occur, albeit at very low efficiency.

CC 15-7 (Immunochemistry)

- IT Transplant and Transplantation
(bone marrow, gene transfer into nonhuman primate **CD34**
-pos. progenitor cells in cell repopulation after)
- IT Transformation, genetic
(of genes, into nonhuman primate **CD34**-pos. progenitor
cells, for cell repopulation after bone marrow transplant)
- IT Hematopoietic precursor cell
(lymphoid, gene transfer into nonhuman primate **CD34**
-pos., cell repopulation after bone marrow transplant of)
- IT Hematopoietic precursor cell
(myeloid, gene transfer into nonhuman primate **CD34**
-pos., cell repopulation after bone marrow transplant of)
- IT **Virus**, animal
(**retro-**, **adenosine deaminase** gene
transfer mediated by, into nonhuman primate **CD34**-pos.
progenitor cells, for cell repopulation after bone marrow
transplant)
- IT Bone marrow
(transplant, gene transfer into nonhuman primate **CD34**
-pos. progenitor cells in cell repopulation after)
- IT 9026-93-1, **Adenosine deaminase**
(gene for human, transfer of, **retrovirus**-mediated, into
nonhuman primate **CD34**-pos. progenitor cells, for cell
repopulation after bone marrow transplant)

L16 ANSWER 2 OF 2 HCA COPYRIGHT 1995 ACS

AN 120:69038 HCA

TI Long-term in vivo expression of a murine **adenosine deaminase** gene in rhesus monkey hematopoietic cells of multiple lineages after **retroviral** mediated gene transfer into **CD34+** bone marrow cells

AU Bodine, David M.; Moritz, Tom; Donahue, Robert E.; Luskey, Barry D.; Kessler, Steven W.; Martin, David I. K.; Orkin, Stuart H.; Nienhuis, Arthur W.; Williams, David A.

CS Clin. Hematol. Branch, Natl. Heart, Lung, Blood Inst., Bethesda, MD, 20892, USA

SO Blood (1993), 82(7), 1975-80

CODEN: BLOOAW; ISSN: 0006-4971

DT Journal

LA English

AB Retroviral mediated gene transfer into stem cells has been proposed as therapy for many inherited hematopoietic diseases. Deficiency of the enzyme adenosine deaminase (ADA) results in depletion of T lymphocytes, causing severe combined immunodeficiency syndrome (SCIDS). In this report, the authors describe retroviral mediated gene transfer of a murine ADA cDNA into Rhesus monkey hematopoietic stem cells. Immunoselected **CD34+** bone marrow cells were exposed to medium contg. the ADA retrovirus during culture on a stromal cell line engineered to express the transmembrane form of stem cell factor. After infusion of autologous, transduced cells into irradiated recipients, gene transfer was obsd. in all three monkeys. The ADA provirus was detected in 2% of circulating granulocytes and T cells from 100 days post-transplantation to longer than 1 yr and in B cells from 250 days post-transplantation and beyond. Mouse ADA activity was detected in peripheral blood cells at approx. 3% the

activity of monkey ADA. Thus, the authors have shown gene transfer into repopulating cells that contribute to all hematopoietic lineages with persistent gene expression. These data provide support for the use of stem cell targeted gene transfer for therapy of ADA deficiency.

CC 1-7 (Pharmacology)

Section cross-reference(s): 3

ST ADA deficiency gene therapy **retrovirus** vector;
hematopoietic stem cell ADA gene transfer; **adenosine deaminase** deficiency gene therapy

IT Hematopoietic precursor cell
(**adenosine deaminase** gene of rats transfer into Rhesus monkey, **retro virus**-mediated, gene therapy of severe combined immunodeficiency syndrome in relation to)

IT Gene, animal
(for **adenosine deaminase**, **retro virus**-mediated transfer of murine, to Rhesus monkey hematopoietic cells, gene therapy of severe combined immunodeficiency syndrome in relation to)

IT Transduction, genetic
(of **adenosine deaminase** gene of rats into Rhesus monkey hematopoietic cells)

IT Genetic vectors
(**retroviral**, for rat **adenosine deaminase** gene transfer into rhesus monkey CD34 + bone marrow cells, enzyme gene expression in hematopoietic cells in relation to)

IT Lymphocyte
(T-cell, **adenosine deaminase** of, after murine gene transfer into Rhesus monkey hematopoietic cells)

IT Therapeutics
(geno-, for **adenosine deaminase** deficiency and severe combined immunodeficiency syndrome, **retro virus**-mediated of murine **adenosine deaminase** gene transfer into Rhesus monkey hematopoietic cells in)

IT Immunodeficiency
(severe combined, gene therapy for, **adenosine deaminase** gene transfer into hematopoietic cells in relation to)

IT 9026-93-1, **Adenosine deaminase**
(gene for, **retro virus**-mediated transfer of murine to Rhesus monkey hematopoietic cells, severe combined immunodeficiency syndrome gene therapy in relation to)

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E KOHN D/AU
L19 224 S E3-9
E BLAESE M/AU
L20 20 S E3-4
E MULLEN C/AU
L21 43 S E3-7
E MOEN R/AU
L22 82 S E3-6
L23 359 S L19 OR L20 OR L21 OR L22
L24 1983 S (RETROVIR? OR RETRO (2W) VIR##) (3A) VECTOR#
L25 5021 S ADENOSINE DEAMINASE
L26 1874 S CD34
L27 88 S L23 AND (L24 OR L25 OR L26)
L28 66 S L23 AND L24
L29 8 S L28 AND L25
L30 1 S L29 AND L26
L31 196 S SEVERE COMBINED IMMUNE DEFICIENCY
L32 3 S L31 AND L23
L33 3 S L24 AND L25 AND L26
L34 0 S L33 AND L31
L35 2237 S SEVERE COMBINED IMMUNODEFICIENCY
L36 16 S L35 AND L23
L37 1 S L35 AND L33
L38 6 S L32 OR L33
L39 1 S L37 OR L30
L40 5 S L38 NOT L39

FILE 'HCA' ENTERED AT 08:48:45 ON 10 JUL 95

FILE 'BIOSIS' ENTERED AT 08:50:01 ON 10 JUL 95

=> d bib ab st l39;d bib ab st l40 1-5

L39 ANSWER 1 OF 1 BIOSIS COPYRIGHT 1995 BIOSIS

AN 94:32442 BIOSIS

DN 97045442

TI Strategies for gene therapy.

AU Blaese R M; Mullen C A; Ramsey W J

CS Cellular Immunol. Sect., Metabolism Branch, National Cancer Inst.,
National Inst. Health, Build. 10, Room 6B05, Bethesda, MD 20892, USA

SO Pathologie Biologie 41 (8). 1993. 672-676. ISSN: 0369-8114

LA English

AB The use of retroviral-mediated gene transfer to introduce a DNA label into T cells (TIL) being used in the immunotherapy of patients with malignant melanoma filially opened the door to the clinical

application of gene therapy for a wide variety of inherited and acquired diseases. The gene therapy trial for ADA deficiency SCID has demonstrated that long-term stable expression of exogenous genes can be achieved in human T lymphocytes using **retroviral vectors** for ex vivo treatment and that significant immune reconstitution can be achieved in these patients following periodic infusions with ADA gene-corrected autologous T cells. Newer clinical applications include the insertion of genes into **CD34** enriched stem cell populations, the testing of autologous tumor vaccines employing cytokine acne-modified tumor cells and the direct transfer of the herpes thymidine kinase gene into brain tumors in situ in order to render those tumors sensitive to treatment with the ordinarily non-cytotoxic drug ganciclovir.

ST JOURNAL ARTICLE; HUMAN T-CELLS; RETROVIRAL-MEDIATED GENE TRANSFER; **SEVERE COMBINED IMMUNODEFICIENCY**; **ADENOSINE DEAMINASE**; CANCER TREATMENT

L40 ANSWER 1 OF 5 BIOSIS COPYRIGHT 1995 BIOSIS

AN 95:236476 BIOSIS

DN 98250776

TI Correction of IL-4 receptor function in lymphoblastoid cell lines (LCL) from patients with X-linked **severe combined immune deficiency** (X-SCID) by retroviral mediated transfer of the gamma-C gene.

AU Uribe L; Taylor N; Smith S; Hong Y-H; Yu X-J; **Kohn D**; Weinberg K

CS Children's Hosp. Los Angeles, Los Angeles, CA, USA

SO 105th Annual Meeting of the American Pediatric Society and the 64th Annual Meeting of the Society for Pediatric Research, San Diego, California, USA, May 7-11, 1995. Pediatric Research 37 (4 PART 2). 1994. 11A. ISSN: 0031-3998

DT Conference

LA English

ST MEETING ABSTRACT; MEETING POSTER; INTERLEUKIN 4; GMMA-C GENE THERAPY

L40 ANSWER 2 OF 5 BIOSIS COPYRIGHT 1995 BIOSIS

AN 93:458859 BIOSIS

DN BA96:103759

TI T LYMPHOCYTE ONTOGENY IN ADENOSINE DEAMINASE-DEFICIENT **SEVERE COMBINED IMMUNE DEFICIENCY** AFTER TREATMENT WITH POLYETHYLENE GLYCOL-MODIFIED ADENOSINE DEAMINASE.

AU WEINBERG K; HERSHFIELD M S; BASTIAN J; **KOHN D**; SENDER L; PARKMAN R; LENARSKY C

CS DIV. RESEARCH IMMUNOL./BONE MARROW TRANSPLANTATION, CHILDRENS HOSPITAL LOS ANGELES, 4650 SUNSET BLVD. 62, LOS ANGELES, CA 90027, USA.

SO J CLIN INVEST 92 (2). 1993. 596-602. CODEN: JCINAO ISSN: 0021-9738

LA English

AB Adenosine deaminase (ADA) deficiency causes **severe combined immune deficiency** (SCID) by interfering with the metabolism of deoxyadenosine, which is toxic to T lymphocytes at all stages of differentiation. Enzyme replacement with polyethylene glycol-modified ADA (PEG-ADA) has been previously

population of pluripotent hematopoietic stem cells capable of self-renewal. We present evidence for the highly efficient gene transfer and sustained expression of human ADA in human primitive hematopoietic progenitors using retroviral supernatant with a supportive stromal layer. A stem cell-enriched (CD34+) fraction was also successfully transduced. Duchenne muscular dystrophy (DMD) is also a good model for somatic gene therapy. Two of the challenges presented by this model are the large size of the gene and the large number of target cells. Germline gene transfer and correction of the phenotype has been demonstrated in transgenic mdx mice using both a full-length and a truncated form of the dystrophin cDNA. We present here a deletion mutagenesis strategy to truncate the dystrophin cDNA such that it can be accommodated by retroviral and adenoviral vectors useful for somatic gene therapy.

ST HUMAN RETROVIRUS COMPLEMENTARY DNA ADENOSINE DEAMINASE PLURIPOTENT HEMATOPOIETIC STEM CELLS DUCHENNE MUSCULAR DYSTROPHY GENETIC ENGINEERING GENE THERAPY THERAPEUTIC METHOD

L40 ANSWER 5 OF 5 BIOSIS COPYRIGHT 1995 BIOSIS

AN 89:279851 BIOSIS

DN BR37:4848

TI SEVERE COMBINED IMMUNE

DEFICIENCY SCID DUE TO DEFECTIVE INTERLEUKIN 2 RECEPTOR ALPHA IL2-R-A EXPRESSION.

AU WEINBERG K I; PARR T; ANNETT G M; COATES T; KOHN D B; LENARSKY C; PARKMAN R

CS DIV. RES. IMMUNOL. BONE MARROW TRANSPLANT., CHILD HOSP. LOS ANGELES, UNIV. SOUTH. CALIF., LOS ANGELES, CALIF., USA.

SO JOINT MEETING OF THE AMERICAN PEDIATRIC SOCIETY AND THE SOCIETY FOR PEDIATRIC RESEARCH, WASHINGTON, D.C., USA, MAY 1-4, 1989. PEDIATR RES 25 (4 PART 2). 1989. 170A. CODEN: PEREBL ISSN: 0031-3998

DT Conference

LA English

ST ABSTRACT HUMAN B LYMPHOCYTE T LYMPHOCYTE SIGNAL TRANSDUCTION

=> fil medlins

'MEDLINS' IS NOT A VALID FILE NAME

SESSION CONTINUES IN FILE 'BIOSIS'

=> fil medl

FILE 'MEDLINE' ENTERED AT 08:50:47 ON 10 JUL 95

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FILE 'BIOSIS' ENTERED AT 08:50:01 ON 10 JUL 95

FILE 'MEDLINE' ENTERED AT 08:50:47 ON 10 JUL 95

L41 1409 S CD34
 E ADENOSINE DEAMINASE/CT
 L42 3191 S (ADENOSINE DEAMINASE+NT)/CT
 E RETROVIRUS/CT
 E RETROVIRIDAE/CT
 E E3+ALL
 E VECTORS/CT
 E E3+ALL
 E VECTORS/CT
 E E8+ALL
 E GENETIC VECTORS+NT/CT
 E GENETIC VECTORS+ALL/CT
 L43 7 S L41 AND L42
 L44 375 S SEVERE COMBINED IMMUNODEFICIENCY+NT/CT
 L45 67 S L44 AND L42 AND L42
 L46 12 S L45 AND (VECTOR# OR RETROVIR?)
 L47 2 S L46 AND L41

=> d bib ab ct 1-2

L47 ANSWER 1 OF 2 MEDLINE *photocopy - milne*
 AN 94002371 MEDLINE
 TI Treatment of severe combined immunodeficiency disease (SCID) due to
 adenosine deaminase deficiency with CD34+ selected
 autologous peripheral blood cells transduced with a human ADA gene.
 Amendment to clinical research project, Project 90-C-195, January
 10, 1992.
 AU Blaese R M; Culver K W; Chang L; Anderson W F; Mullen C; Nienhuis A;
 Carter C; Dunbar C; Leitman S; Berger M; et al
 SO Hum Gene Ther, (1993 Aug) 4 (4) 521-7.
 Journal code: A12. ISSN: 1043-0342.
 CY United States
 DT Journal; Article; (JOURNAL ARTICLE)
 LA English
 FS Priority Journals
 EM 9401
 AB Significant increases in lymphocyte adenosine deaminase activity, T
 cell numbers and immune function have been achieved in the two
 children with SCID thus far treated with autologous T cells
 genetically-corrected by retroviral-mediated insertion of
 a normal ADA gene. Although the data obtained to date demonstrate
 that the use of ADA gene corrected peripheral T cells appears to be
 an effective treatment for ADA(-)SCID, it is theoretically
 preferable to try to develop a treatment for these children that
 will result in stem cell gene correction. The genetic correction of
 T cell progenitors with long-term immune reconstituting ability
 would be more desirable because repeated infusions of genetically
 altered cells should not be necessary and the generation of a more
 complete repertoire of T cell specificities might also be possible.

Furthermore, the present treatment protocol involves indefinite continuation of enzyme replacement treatment with PEG-ADA. The demonstration of ADA gene expression in the progeny of transduced stem cells may simplify the decision concerning cessation of this very costly enzyme treatment (approximately \$250,000/yr./patient). Recent evidence suggests that a small fraction of bone marrow or peripheral blood mononuclear cells bearing the **CD34** antigen contains hematopoietic stem cells with both lymphoid and myeloid reconstituting ability. We propose in this amendment to supplement the infusion of human ADA gene-transduced autologous T cells in children with ADA(-)SCID with autologous peripheral blood **CD34+** cells transduced with a second, readily distinguishable ADA vector. (ABSTRACT TRUNCATED AT 250 WORDS)

CT Check Tags: Human
Adenosine Deaminase: DF, deficiency
***Adenosine Deaminase: GE, genetics**
***Antigens, CD**
 Cells, Cultured
 Clinical Protocols
***Gene Therapy**
***Hematopoietic Stem Cell Transplantation**
 Hematopoietic Stem Cells: IM, immunology
 Hematopoietic Stem Cells: ME, metabolism
***Hematopoietic Stem Cells: TR, transplantation**
Severe Combined Immunodeficiency: EN, enzymology
Severe Combined Immunodeficiency: GE, genetics
***Severe Combined Immunodeficiency: TH, therapy**
 T-Lymphocytes: IM, immunology
 T-Lymphocytes: TR, transplantation
 Transduction, Genetic
 Transplantation, Autologous

L47 ANSWER 2 OF 2 MEDLINE
 AN 93226692 MEDLINE
 TI Gene transfer therapy for heritable disease: cell and expression targeting.
 AU Mitani K; Clemens P R; Moseley A B; Caskey C T
 CS Howard Hughes Medical Institute, Baylor College of Medicine, Houston, Texas 77030.
 NC R01 DK42696 (NIDDK)
 SO Philos Trans R Soc Lond B Biol Sci, (1993 Feb 27) 339 (1288) 217-24.
 Ref: 70
 Journal code: P5Z. ISSN: 0080-4622.
 CY ENGLAND: United Kingdom
 DT Journal; Article; (JOURNAL ARTICLE)
 General Review; (REVIEW)
 (REVIEW, TUTORIAL)
 LA English
 FS Priority Journals
 EM 9307
 AB Gene therapy is defined as the delivery of a functional gene for expression in somatic tissues with the intent to cure a disease. Different gene transfer strategies may be required to target different tissues. Adenosine deaminase (ADA) deficiency is a good

gene therapy model for targeting a rare population of pluripotent hematopoietic stem cells capable of self-renewal. We present evidence for the highly efficient gene transfer and sustained expression of human ADA in human primitive hematopoietic progenitors using **retroviral** supernatant with a supportive stromal layer. A stem cell-enriched (CD34+) fraction was also successfully transduced. Duchenne muscular dystrophy (DMD) is also a good model for somatic gene therapy. Two of the challenges presented by this model are the large size of the gene and the large number of target cells. Germline gene transfer and correction of the phenotype has been demonstrated in transgenic mdx mice using both a full-length and a truncated form of the dystrophin cDNA. We present here a deletion mutagenesis strategy to truncate the dystrophin cDNA such that it can be accommodated by **retroviral** and **adenoviral vectors** useful for somatic gene therapy.

CT Check Tags: Human; Support, Non-U.S. Gov't; Support, U.S. Gov't, P.H.S.

Adenosine Deaminase: DF, deficiency

Adenosine Deaminase: GE, genetics

Dystrophin: GE, genetics

*Gene Therapy: MT, methods

Genetic Vectors

*Hereditary Diseases: TH, therapy

Muscular Dystrophy: TH, therapy

Severe Combined Immunodeficiency: TH, therapy

Transfection: MT, methods